

Image by Kara Ruff, 2019

# Pathogens or symbionts? A study of the slime mold *Cavenderia aureostipes* var. *Helvetia* and its associated bacteria

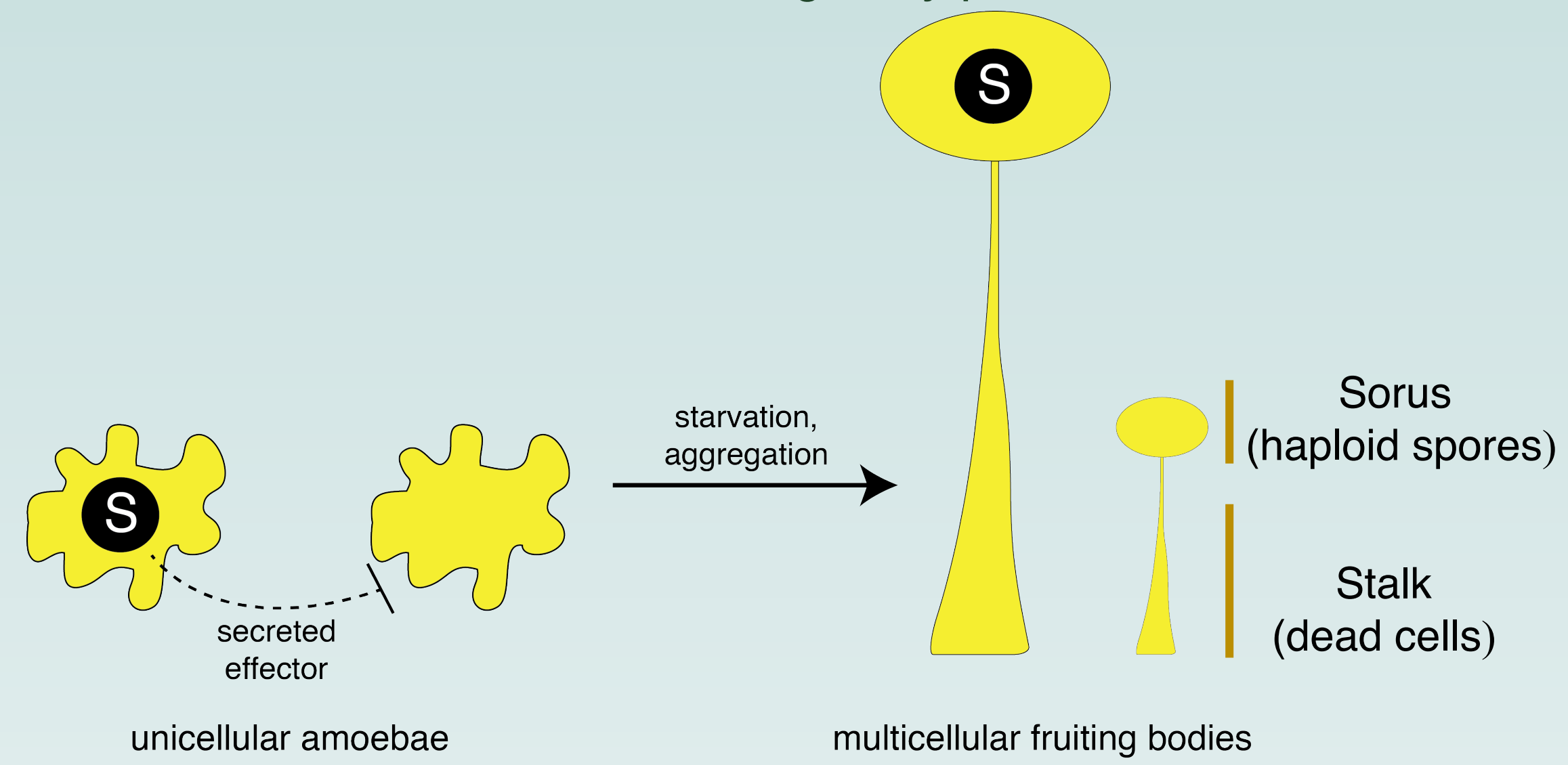
Kara Ruff - Department of Biology, University of Victoria, Victoria, BC  
Supervised by Dr. Ryan Gawryluk



University of Victoria

## INTRODUCTION

- Studies on slime molds have found an intracellular bacterial symbiont able to persist through the “sterile” stalk and fruiting body stage<sup>1</sup>
- Burkholderia* sp. confer the ability of *D. discoideum* to “farm” food bacteria<sup>2,3</sup> and secrete an effector that harms the fruiting body production of non-farmer amoeba<sup>4</sup>



**Figure 1.** The cellular slime mole *Dictyostelium discoideum* has an intracellular symbiont (*Burkholderia* sp.) that secretes an effector molecule harming fruiting body production of “non-farming” amoeba. The symbiont can persist through multiple social cycles in the stalk and spores of *D. discoideum*.

- We surveyed wild slime mold isolates for possible symbionts
- We found 3 culturable bacterial species associated with *C. aureostipes* var. *Helvetia* that were somehow able to persist through multiple social cycles:
  - Achromobacter* species 1
  - Achromobacter* species 2
  - Escherichia coli* strain 1

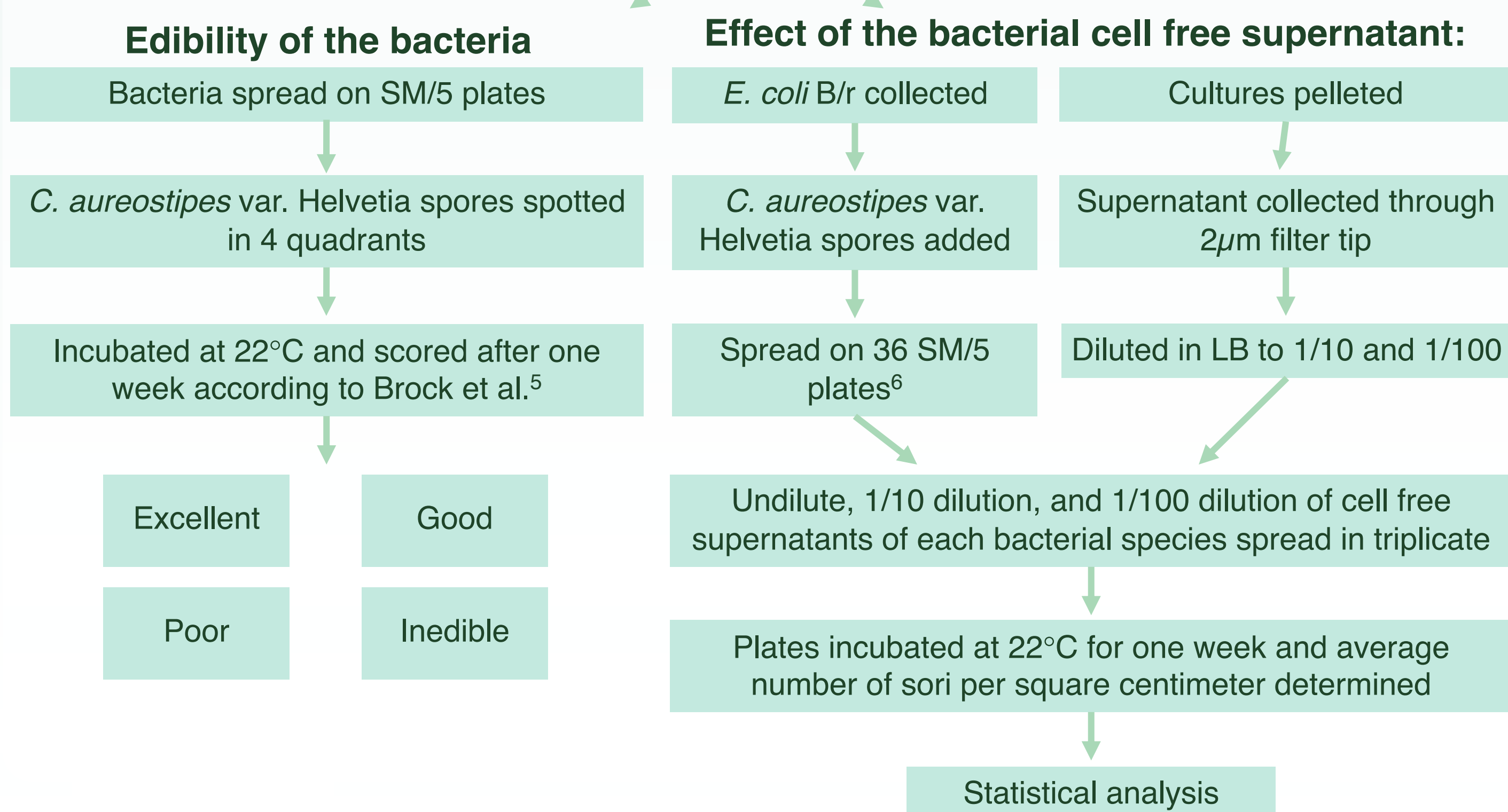
## OBJECTIVES

To investigate the relationship between *C. aureostipes* var. *Helvetia* and associated bacteria by examining:

- location of the bacteria (**intracellular** or **extracellular**?)
- edibility of the bacteria (**edible** or **inedible**?)
- effect of the bacterial cell free supernatant (**promote** or **inhibit** growth?)

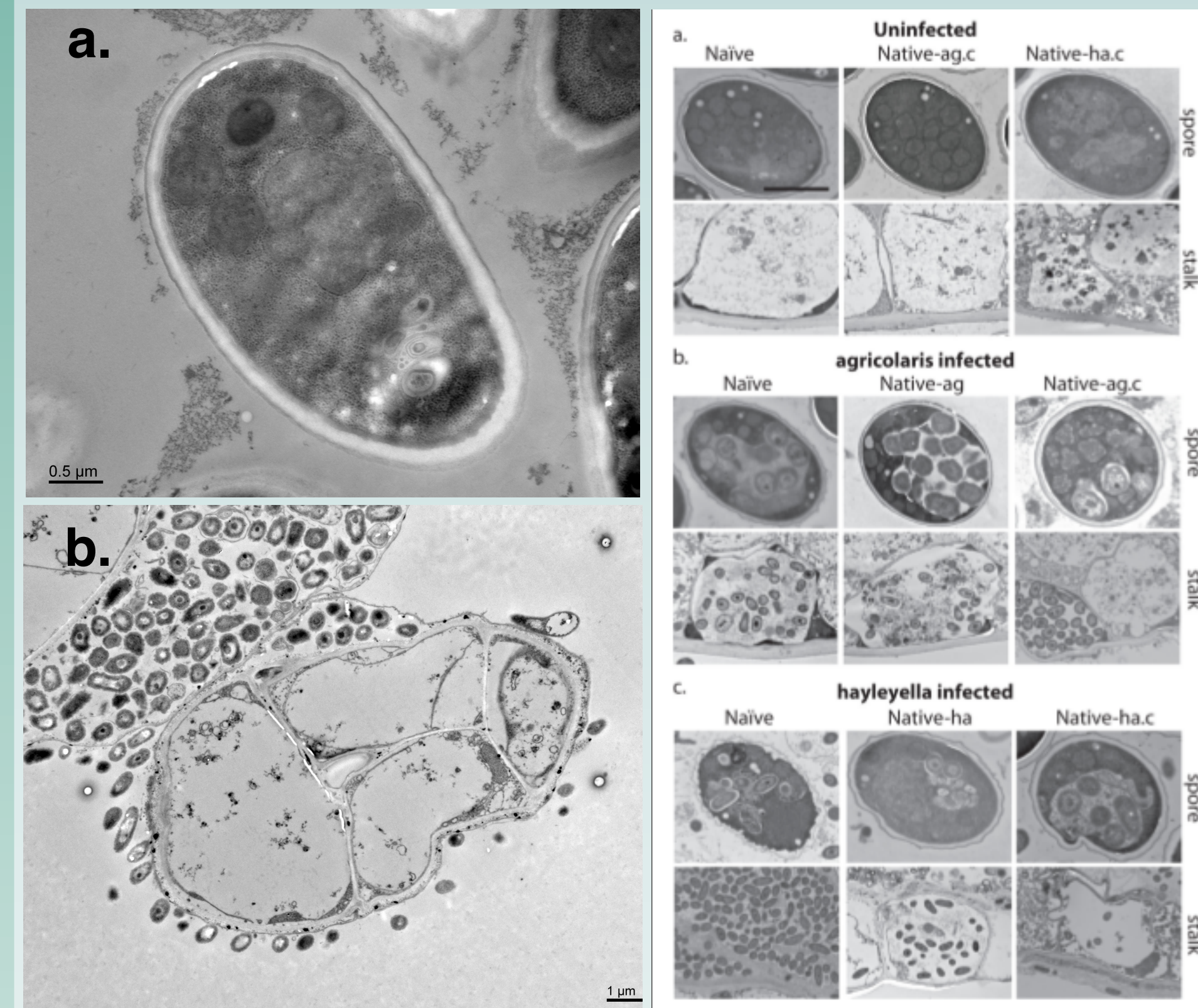
## METHODS

*E. coli* B/r, *E. coli* Strain 1 *Achromobacter* sp. 1 and sp. 2 cultures grown at 28°C with shaking at 150rpm



## RESULTS

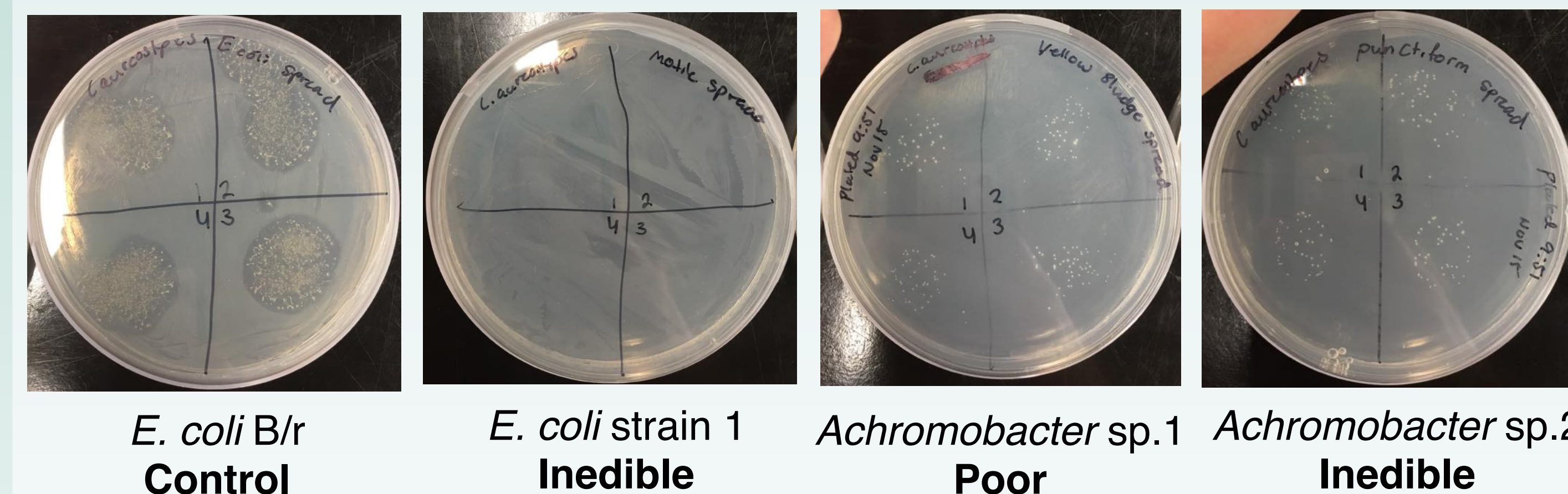
### Location of the bacteria:



- No intracellular infection of spores or stalk
- Appearance of bacterial aggregates in a possible biofilm suggests bacteria may be maintained through multiple social cycles simply by “hanging on” to sori

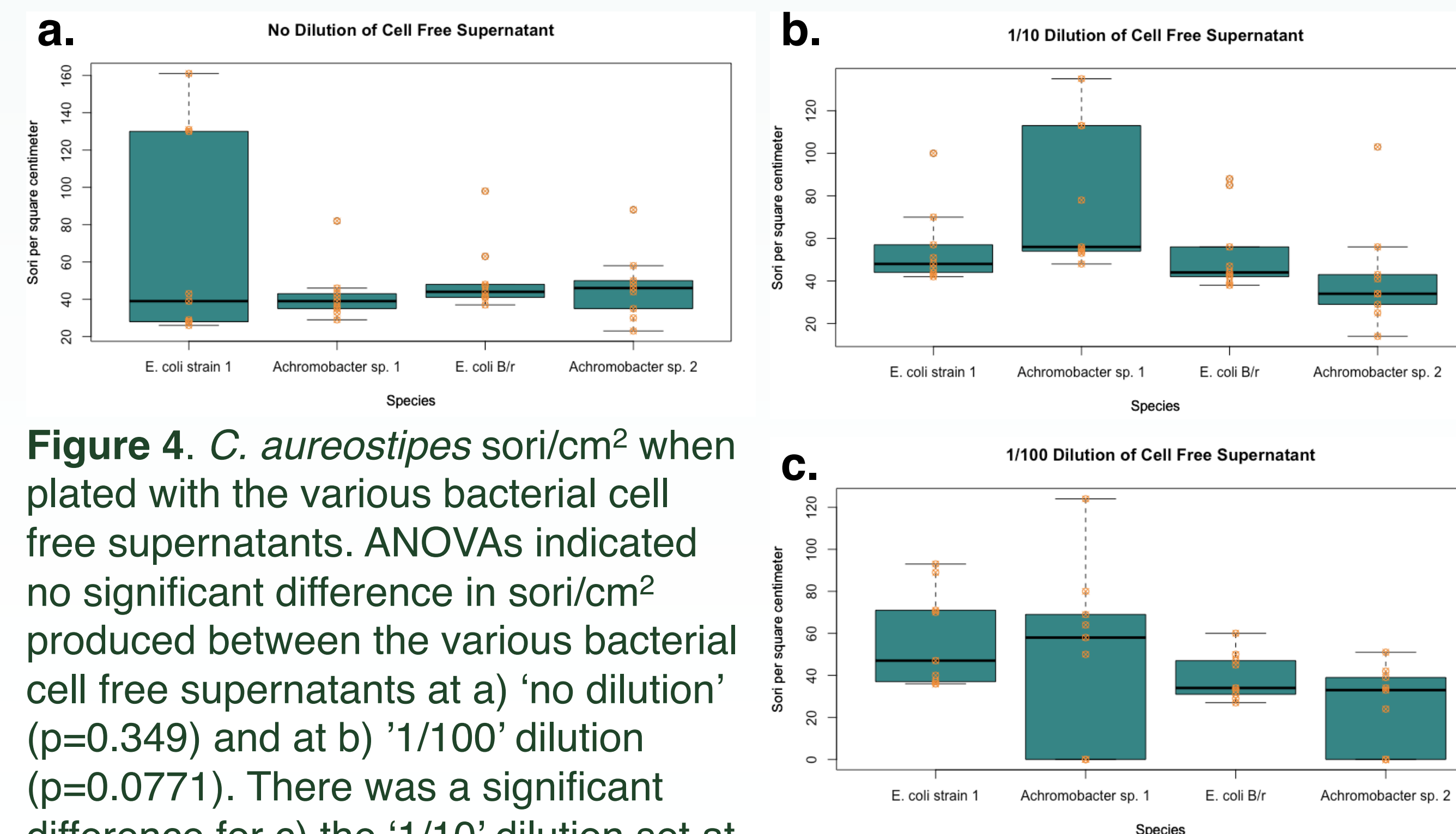
**Figure 2.** Comparison of TEM images of a) spores and b) stalk cells to Figure 6 by Shu et al.<sup>1</sup>

### Edibility of the bacteria:



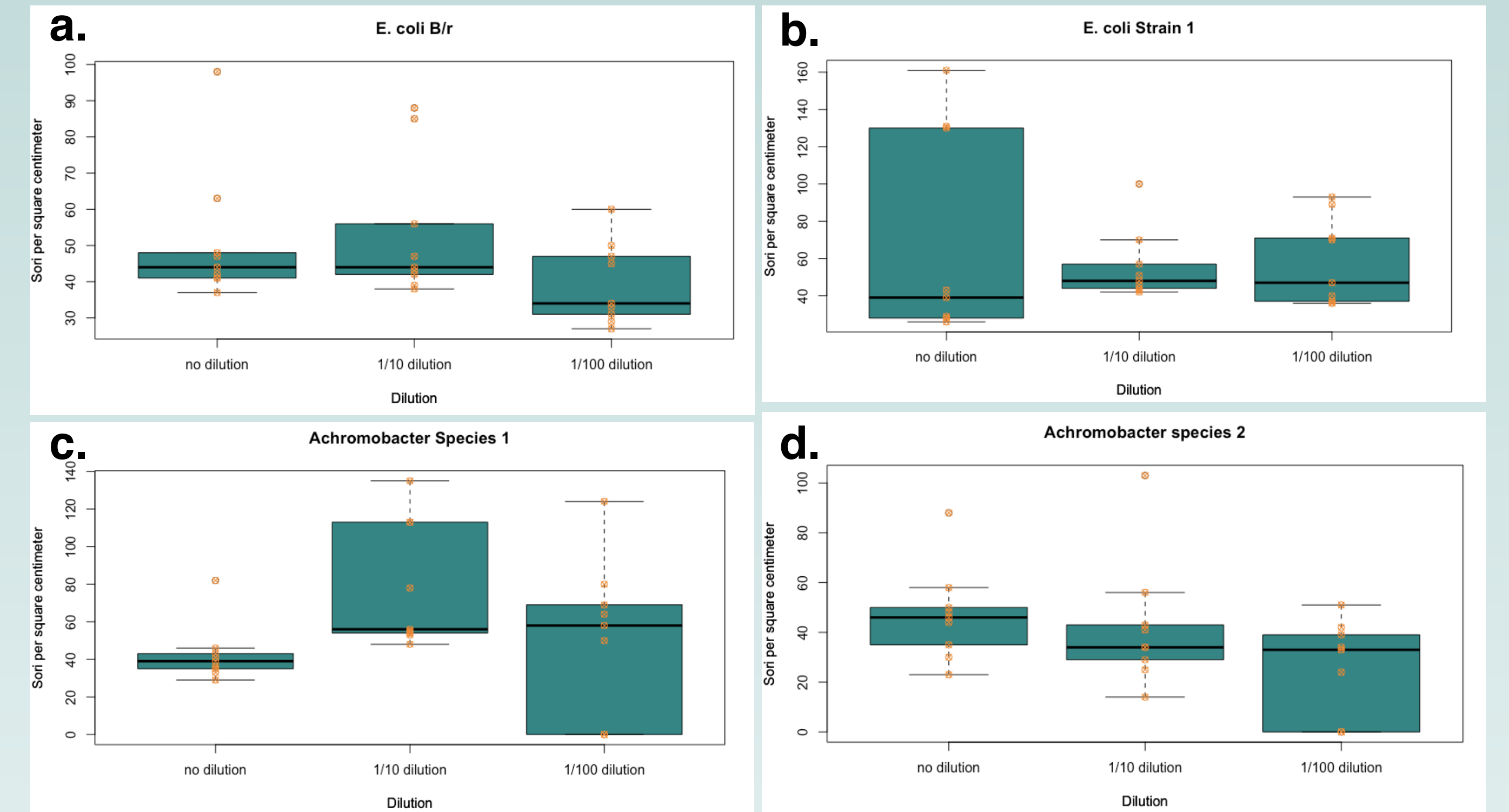
**Figure 3.** Co-cultured bacteria are poorly edible in comparison to food bacteria (*E. coli* B/r). *Achromobacter* inedibility may be due to biofilm formation<sup>7</sup>. *E. coli* str. 1 may be pathogenic toward *C. aureostipes* based on observations and previous reports<sup>8,9</sup>.

### Effect of the bacterial cell free supernatant (between species):



**Figure 4.** *C. aureostipes* sori/cm<sup>2</sup> when plated with the various bacterial cell free supernatants. ANOVAs indicated no significant difference in sori/cm<sup>2</sup> produced between the various bacterial cell free supernatants at a) ‘no dilution’ (p=0.349) and at b) ‘1/100’ dilution (p=0.0771). There was a significant difference for c) the ‘1/10’ dilution set at p=0.032. F-test indicated that difference was due to *Achromobacter* sp. 1 (p=0.046).

### Effect of the bacterial cell free supernatant (within species):



**Figure 5.** No dosage-dependent effect of co-cultured bacteria on *C. aureostipes* fruiting body density. Sori/cm<sup>2</sup> plated with cell free supernatant from a) *E. coli* B/r (p=0.195), b) *E. coli* Strain 1 (p=0.731), c) *Achromobacter* sp. 1 (p=0.065), and d) *Achromobacter* sp. 2 (p=0.096).

## CONCLUSIONS

- The bacteria are **NOT** intracellular symbionts of *C. aureostipes*
- Achromobacter* sp. 1 has poor edibility and *Achromobacter* sp. 2 is inedible, while *E. coli* species 1 may be pathogenic
- Significant reduction in sori/cm<sup>2</sup> for the 1/10 dilution of *Achromobacter* sp. 2 likely due to human error: dose-dependent effect not observed
- Analysis into the genetic basis for inedibility/resistance may provide insight into the possibly pathogenic basis of these bacteria for both Dictyostelids

## REFERENCES

- Shu L, et al. 2018. Symbiont location, host fitness, and possible coadaptation in a symbiosis between social amoebae and bacteria. eLife 7:e42660. DOI: <https://doi.org/10.7554/eLife.42660>
- Brock DA, et al. 2011. Primitive agriculture in a social amoeba. Nature 469:393.
- DiSalvo S, et al. 2015. Burkholderia bacteria infectiously induce the proto-farming symbiosis of Dictyostelium amoebae and food bacteria. Proc Natl Acad Sci U S A 112:E5029-E5037.
- Brock DA, Jones K, Queller DC, Strassmann JE. 2016. Which phenotypic traits of Dictyostelium discoideum farmers are conferred by their bacterial symbionts? Symbiosis 68:39-48.
- Brock DA, et al. 2018. Diversity of free-living environmental bacteria and their interactions with a bacterivorous amoeba. Front Cell Infect Microbiol 8:12.
- dictyBase. Media and Buffers. <http://dictybase.org/techniques/media/media.html#SM5>
- Nielsen SM, Penstoft LN, Nørskov-Lauritsen, N. 2019. Motility, Biofilm formation and antimicrobial efflux of sessile and planktonic cells of Achromobacter xylosoxidans. Pathogens 8 (1):14. doi: 10.3390/pathogens8010014.
- Adiba S, et al. 2010. From grazing resistance to pathogenesis: the coincidental evolution of virulence factors. PLoS ONE 5(8): e11882. <https://doi.org/10.1371/journal.pone.0011882>
- Snyder M, et al. 2019. Identification and characterization of Escherichia coli genes associated with grazing resistance to the social amoeba Dictyostelium discoideum. J Immunol.

## ACKNOWLEDGEMENTS

This research was supported by the Jamie Cassels Undergraduate Research Award at the University of Victoria. I would like to thank Dr. Ryan Gawryluk for allowing me to use his resources and laboratory to conduct the research, and his oversight and guidance during the investigation.